

TWO GUAIANOLIDES FROM *CALEA SOLIDAGINEA*

ALFONSO G. OBER, LOWELL E. URBATSCH* and NIKOLAUS H. FISCHER†

Department of Chemistry and *Department of Botany, Louisiana State University, Baton Rouge, LA 70803, U.S.A.

(Revised received 20 March 1985)

Key Word Index—*Calea solidaginea*; Asteraceae; Heliantheae; sesquiterpene lactones; guaianolides; solidaginolides A and B.

Abstract—The isolation and structure elucidation of two new guaianolides, solidaginolides A (1) and B (5), from *Calea solidaginea* are reported. The structures of the new compounds were established by spectroscopic methods.

INTRODUCTION

Among the 40 *Calea* species that have so far been chemically investigated [1], guaianolides were found only in *C. subcordata* [2], *C. berteriana* [1], *C. prunifolia* [1] and *C. solidaginea* [1]. In continuation of our biochemical systematic investigations of the genus *Calea* we have further analysed *C. solidaginea* from Venezuela for its sesquiterpene lactone constituents. Previous studies of the same collection of this species [1] afforded the two guaianolides 8-epi-8-tiglylrupicolin A and desacyl-8-tiglylsolidaginatolide A. We now report the isolation and structure determination of two new guaianolides, solidaginolide A (1) and solidaginolide B (5). The structures of the new compounds were established by spectroscopic and chemical methods.

RESULTS AND DISCUSSION

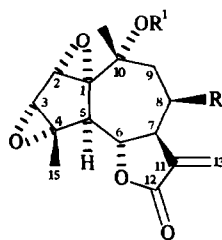
Solidaginolide A (1), $C_{20}H_{24}O_7$, is a gum with an IR spectrum showing the presence of hydroxyl group(s) (3640 and 3600 cm^{-1}), a γ -lactone (1760 cm^{-1}), an unsaturated ester (1700 cm^{-1}), and carbon-carbon double bond(s) (1650 and 1605 cm^{-1}). The ^1H NMR spectrum corroborated the presence of an α -methylene- γ -lactone moiety by exhibiting two one-proton doublets at δ 6.24 (H-13a) and 5.47 (H-13b), and a one-proton multiplet at δ 3.74 (H-7). The unsaturated ester side chain was identified as a tiglate group on the basis of the characteristic ^1H NMR signals (a one-proton quartet of quartets at δ 6.86, and two three-proton vinyl methyl signals at δ 1.76 and 1.79), together with strong mass spectral peaks at m/z 83 (A^1) and 55 (A^2). Detailed spin decoupling experiments in CDCl_3 and benzene- d_6 allowed the assignments of all ^1H NMR signals (Table 1). The mass spectrum of 1 showed no molecular ion peak and only very weak high-mass peaks. The spectrum was dominated by a base peak at m/z 111 which was assigned to $C_6H_7O_2^+$ (B). The formation of this ion can be accounted for by fragmentation of a diepoxycyclopentane moiety, thus suggesting

structural features in 1 similar to canin (4) [3] and artecamin (2) [4].

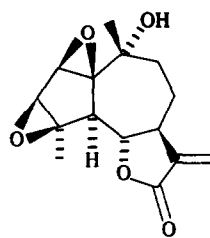
The chemical shifts and multiplicities of H-5 (δ 2.86, d , $J_{5,6} = 10.5\text{ Hz}$) and H-6 (4.72, t , $J_{5,6} = J_{6,7} = 10.5\text{ Hz}$), together with the absence of angular methyl signals in the ^1H NMR spectrum, suggested a guaianolide-type skeleton with a tiglate ester side chain. Assuming that H-7 is α -oriented as in all known lactones from higher plants [5], H-6 should be β -oriented since the coupling constant ($J_{6,7} = 10.5\text{ Hz}$) clearly indicated an antiperiplanar arrangement of H-6 and H-7. Similarly, the coupling constant $J_{5,6} = 10.5\text{ Hz}$ also supported an antiperiplanar arrangement of H-5 and H-6, and therefore an α -orientation for H-5. On the basis of chemical shift arguments, the ester side chain must be attached to C-8. Furthermore, the small coupling constant ($J_{7,8} = 2.5\text{ Hz}$) suggested a β -orientation of the C-8 ester sidechain. The chemical shifts of the methyl signals at δ 1.12 (H-14) and 1.58 (H-15) were in accord with the presence of a hydroxyl group at C-10 and an epoxide function at C-4, respectively. A narrow doublet ($J = 1\text{ Hz}$) at δ 3.60 (H-2) which was coupled to a broadened one-proton singlet at 3.30 (H-3) suggested epoxide functions at C-1 and C-3 as in canin (4) [3] and artecamin (2) [4], further corroborating the mass spectral conclusions. The stereochemistry of the epoxide functions and the orientation of the hydroxyl at C-10 in 1 were assigned by NMR spectral correlations of H-5 and H-6 with those reported for artecamin (2) [4] and 10-epicanin [6]. The chemical shift of H-5 in compound 1 (δ 2.86) is comparable with that of artecamin (2) (2.83), and appears 0.31 ppm and 0.24 ppm downfield from the respective H-5 signals of canin (4) and 10-epicanin [6]. It must therefore be concluded that the epoxide functions at C-1 and C-3 in 1 are α -oriented as in artecamin (2).

In situ acylation of 1 with trichloroacetyl isocyanate [7] yielded the trichloroacetyl carbamate derivative 3, the ^1H NMR spectrum of which showed one NH signal at δ 8.47 which confirmed the presence of one hydroxyl group in compound 1. The assignment of the ^1H NMR signals of 3 were deduced by double irradiation experiments (Table 1). The paramagnetic acylation shift of the C-10 methyl protons (H-14) from δ 1.12 in 1 to 1.40 in the trichloroacetyl carbamate 3 ($\Delta\delta = 0.28$) was in agreement with the presence of a hydroxyl group at C-10. Absence of

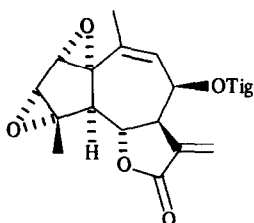
†To whom correspondence should be addressed.



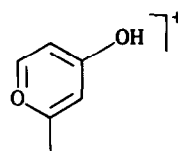
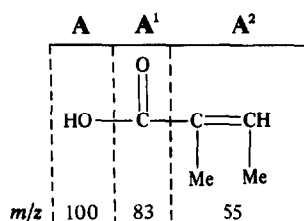
- 1** R = OTiglate, R¹ = H
2 R = H, R¹ = H
3 R = OTiglate, R¹ = CONHCOCCl₃



4



5



B

Table 1. ¹HNMR spectral data* of solidagininolide A (**1**) and B (**5**) and derivative **3** (200 MHz, CDCl₃[C₆D₆], TMS)

H	1	3	5
2	3.60 [2.83] <i>d</i> (1.0)	3.77 <i>br s</i>	3.78 [3.37] <i>br s</i>
3	3.30 [2.62] <i>br s</i>	3.39 <i>br s</i>	3.72 [3.29] <i>br s</i>
5	2.86 [2.58] <i>d</i> (10.5)	2.36 <i>d</i>	2.40 [2.09] <i>d</i> (10.0)
6	4.72 [4.95] <i>dd</i> (10.5; 10.5)	4.75 <i>t</i>	5.01 [5.18] <i>dd</i> (10.0; 10.0)
7	3.74 [3.47] <i>m</i>	3.35 <i>m</i>	4.26 [4.14] <i>m</i>
8	5.65 [5.62] <i>ddd</i> (9.0; 2.5; 2.5)	5.71 <i>dt</i>	6.08 [6.10] <i>m</i> (narrow)
9 α	2.36 [1.98] <i>dd</i> (16.0; 9.0)	2.79 <i>dd</i>	
9 β	1.86 [1.88] <i>dd</i> (16.0; 2.5)	1.92 <i>m</i>	5.48 [5.40] <i>m</i>
13a	6.24 [6.14] <i>d</i> (4.0)	6.28 <i>d</i>	6.30 [6.28] <i>d</i> (4.0)
13b	5.47 [5.20] <i>d</i> (3.3)	5.53 <i>d</i>	5.54 [5.26] <i>d</i> (3.3)
14	1.12 [0.57] <i>s</i>	1.40 <i>s</i>	1.85 [1.52] <i>br s</i>
15	1.58 [1.41] <i>s</i>	1.58 <i>s</i>	1.60 [1.36] <i>s</i>
Tig	6.86 [6.96] <i>qq</i> (7.5; 2.0)	6.88 <i>m</i>	6.78 [6.82] <i>qq</i> (7.5; 2.0)
	1.79 [1.71] <i>br</i>	1.81 <i>br</i>	1.76 [1.69] <i>br</i>
	1.76 [1.67] <i>br</i>	1.80 <i>br</i>	1.75 [1.64] <i>br</i>
NH	—	8.47 <i>s</i>	—

*Coupling constants (*J*) or line separations in Hz are given in parentheses. Multiplets are given by the usual symbols.

a paramagnetic acylation shift of H-6 β in the carbamate derivative 3 (δ 4.75), when compared with H-6 in 1 (4.72), suggested an C-10 α -hydroxyl group. More significantly, the paramagnetic shift of H-5 α and H-7 α was 0.50 and 0.39, respectively. This permitted the unambiguous assignment of the stereochemistry as C-10 α -hydroxyl and an overall stereostructure for solidaginolide A as shown in 1.

Solidaginolide B (5), C₂₀H₂₂O₆, exhibited in the ¹H NMR spectrum two one-proton doublets at δ 6.30 (H-13a) and 5.54 (H-13b), and a one-proton multiplet at 4.26 (H-7) suggesting an α -methylene- γ -lactone. A strong IR band at 1760 cm⁻¹ corroborated the presence of a γ -lactone moiety. Further IR absorption bands at 1700, 1650 and 1605 cm⁻¹ indicated the presence of an unsaturated ester side chain and double bonds. As in the case of compound 1, the ester side chain was identified as a tiglic ester on the basis of the diagnostic ¹H NMR and mass spectral signals. Exhaustive spin decoupling experiments in CDCl₃ and benzene-*d*₆ allowed the assignments of the skeletal protons (Table I).

Comparison of the ¹H NMR spectra of compounds 1 and 5 showed major differences only for the signals of H-9 and H-14. Instead of the H-9 proton absorptions, which in 1 were represented by two one-proton doublet of doublets at δ 2.36 (H-9a) and 1.86 (H-9b), in compound 5 a one-proton multiplet appeared at δ 5.48 clearly indicating its olefinic nature. Furthermore, the H-14 signal which in 1 was a three-proton singlet at δ 1.12, appeared in 5 as a broadened three-proton singlet at δ 1.85 which sharpened upon irradiation of the H-9 signal at 5.48. The downfield shift of H-7 (δ 4.26) in compound 5, when compared to 1 (3.74), can be explained if deshielding of this proton by the carbonyl of the ester group at C-8 is assumed. Inspection of stereomodels revealed that this is the case when the seven-membered ring adopts a conformation with the C-8 ester group being in the more favored equatorial position. Based on the above arguments, we postulated stereostructure 5 for solidaginolide B.

EXPERIMENTAL

Calea solidaginea Kunth was collected on December 12, 1979 in Salom, Venezuela (L. Urbatsch No. 3466, voucher deposited at

L.S.U., U.S.A.). The air-dried plant material (728 g) was extracted and worked up as previously described [8, 9]. For the isolation of minor constituents, portions of fractions 23–24 (51 mg) were rechromatographed on silica gel plates using CHCl₃–Me₂CO (9:1), yielding 8 mg of solidaginolide A (1) and 7 mg of solidaginolide B (5).

Solidaginolide A (1), C₂₀H₂₄O₇, gum; UV λ _{MeOH} nm: strong end absorption; IR ν _{max}^{CHCl₃} cm⁻¹: 3640 (OH), 3600 (OH, broad), 1760 (γ -lactone), 1700 (unsaturated ester), 1650 (double bond), 1605 (double bond); EIMS (probe) (rel. int.): 276 [M – A]⁺ (0.2), 258 [M – A – H₂O]⁺ (0.3), 230 [M – A – H₂O – CO]⁺ (0.5), 202 [M – A – H₂O – 2CO]⁺ (1.0), 111 [B]⁺ (100.0), 83 [A]⁺ (68.6), 55 [A']⁺ (18.4).

Solidaginolide B (5), C₂₀H₂₂O₆, gum; UV λ _{MeOH} nm: strong end absorption; IR ν _{max}^{CHCl₃} cm⁻¹: 1760 (γ -lactone), 1700 (unsaturated ester), 1650 (double bond), 1605 (double bond); EIMS (probe) (rel. int.): 258 [M – A]⁺ (0.1), 240 [M – A – H₂O]⁺ (0.1), 83 [A]⁺ (100.0), 55 [A']⁺ (24.7).

Acknowledgements—We thank Helga D. Fischer for technical assistance. A.G.O. thanks the Universidad Tecnica Santa Maria, Valparaiso, Chile for educational leave.

REFERENCES

1. Ober, A. G. (1984) Ph.D. dissertation, Louisiana State University, Baton Rouge.
2. Ober, A. G., Quijano, L., Urbatsch, L. E. and Fischer, N. H. (1984) *Phytochemistry* **23**, 1289.
3. Lee, K. H., Simpson, R. F. and Geissman, T. A. (1969) *Phytochemistry* **8**, 1515.
4. Bhadane, N. R. and Shafizadeh, F. (1975) *Phytochemistry* **14**, 2651.
5. Fischer, N. H., Olivier, E. J. and Fischer, H. D. (1979) in *Progress in the Chemistry of Organic Natural Products* (Herz, W., Grisebach, H. and Kirby, G. B., eds). Springer, Wien.
6. Bohlmann, F. and Zdero, C. (1982) *Phytochemistry* **21**, 2543.
7. Samek, Z. and Budesinsky, M. (1979) *Coll. Czech. Chem. Commun.* **44**, 558.
8. Fischer, N. H., Wiley, R. A., Lin, H. N., Karimian, K. and Politz, S. M. *Phytochemistry* **14**, 2241.
9. Ober, A. G., Urbatsch, L. E. and Fischer, N. H. (1985) *Phytochemistry* **24**, 795.